



# Synthesis and antiplasmodial assessment of nitazoxanide and analogs as new antimalarial candidates

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## Abstract

During the last years, the progression to control malaria disease seems to be slowed and WHO (World Health Organization) reported a modeling analysis with the prediction of the increase in malaria morbidity and mortality in sub-Saharan Africa during the COVID-19 pandemic. A rapid way to the discovery of new drugs could be carried out by performing investigations to identify drugs based on repurposing of “old” drugs. The 5-nitrothiazole drug, Nitazoxanide was shown to be active against intestinal protozoa, human helminths, anaerobic bacteria, viruses, etc. In this work, Nitazoxanide and analogs were prepared using two methodologies and evaluated against *P. falciparum* 3D7. A bithiazole analog, showed attractive inhibitory activity with an EC<sub>50</sub> value of 5.9 μM, low propensity to show toxic effect against HepG2 cells at 25 μM, and no cross-resistance with standard antimalarials.

**Keywords** Nitazoxanide · Analogues · Thiazoles · Malaria

## Introduction

Malaria is caused by *Plasmodium* spp. parasites and transmitted to human by the *Anopheles* vector mosquito. The WHO 2020 report, estimated 229 million malaria cases in 2019, principally in the sub-Saharan region [1]. During the last years, the progression to control the disease seems to be slowed as the estimated deaths in 2019 were 409,000, compared with 405,000 deaths in 2018 and 585,000 in

2010, with 67% of infants under-five. The COVID-19 pandemic has impacted on the investigation and control of infectious diseases [2–4]. As a result of 75% reduction in routine malaria control measures, WHO (World Health Organization) reported a modeling analysis with the prediction of the increase in malaria morbidity and mortality in sub-Saharan Africa during the COVID-19 pandemic [5]. Moreover, the emergence of *Anopheles* vector resistance to the currently used insecticides and of malaria parasite resistance to the available drugs, including artemisinin and its derivatives [6–8], jeopardize malaria control achievements. Thus, there is an urgent need of new and safe drugs to treat this illness.

It is well known that drug discovery and development take about 12–15 years and the process presents a substantial cost. However, an attractive and rapid way to the discovery of new drugs could be carried out by performing investigations to identify drugs to treat rare or common diseases based on repurposing of “old” drugs [9, 10]. Thiazole is an important class of five-membered heterocyclic compounds with diverse biological properties, thus it is found in many drugs such as sulfathiazol (antimicrobial drug), ritonavir (antiretroviral drug), abafungin (antifungal drug), and tiaozofurin (antineoplastic drug) [11].

The 5-nitrothiazole drug, nitazoxanide (Ntz, 1 Fig. 1) presents various biological activities. Ntz was first

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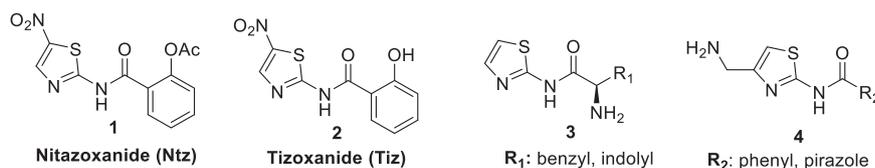
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**Fig. 1** Antiplasmodial heterocycles: nitazoxanide (1), tizoxanide (2), thiazole derived amino acids (3), and aminomethylthiazole carboxamide derivatives (4)



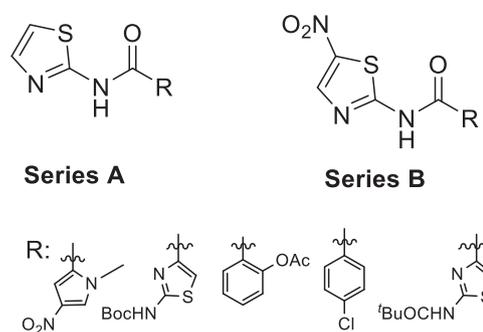
synthesized by Rossignol et al. [12, 13], in the early 1970s as a veterinary anthelmintic agent. Then, this drug was shown to be active against intestinal protozoa, human helminths, and anaerobic bacteria [14–16]. In humans, Ntz is rapidly metabolized to tizoxanide (Tiz), which is as effective against these parasites and bacteria as the parent drug (Fig. 1). The initial indication for nitazoxanide was for cryptosporidiosis and giardiasis, but subsequent investigations have demonstrated that the drug is effective for amoebiasis [17]. In addition, Ntz is active against non-protozoan parasites, including the intestinal tapeworm *Hymenolepis nana* [18], and *Trichuris trichiura* [19]. Moreover, many researchers concluded that it presents antiviral activity against enteric viruses, [20–22] MERS virus [23], and that Ntz and analogs are promising compounds to treat hepatitis C infection [24–26]. Furthermore, several investigations were focused on the mechanisms of action [27–30].

During the last year, vast investigations to find drugs repositioning efficient against SARS-CoV2 were performed. As part of these studies, Ntz has demonstrated in vitro antiviral activity against SARS-CoV2 [30, 31]. In addition, it was suggested that this drug amplifies host innate immune antiviral responses by triggering foreign cytoplasmic RNA sensing and the type 1 interferon axis [32].

Navarrete-Vazquez et al. [33] synthesized two benzologues of Ntz and Tiz and compared their in vitro antiparasitic activities against *Giardia intestinalis*, *Entamoeba histolytica*, *Trichomonas vaginalis*, *Trypanosoma cruzi*, *Leishmania mexicana* and the rodent malaria parasite *Plasmodium berghei*. Ntz and Tiz showed  $IC_{50}$  values in the low micromolar range against *P. berghei*, ( $IC_{50}$ s of 3.9  $\mu$ M and 5.2  $\mu$ M, respectively). Furthermore, in a recent review, Sharma and Prashe highlighted several thiazole derived amino acids (3) [34], and aminomethylthiazole carboxamide derivatives (4) (Fig. 1) [35], as promising anti-malarials with  $EC_{50}$  values in the low micromolar or submicromolar range, respectively [36]. Considering these results, we decided to explore the “old” and versatile drug Ntz, its metabolite Tiz and bis-heterocycles analogs as part of a research program for the synthesis of potential anti-malarials [37–47].

In this work, we present the synthesis and the antiplasmodial activity of Ntz, Tiz, and 2-aminothiazole analogs (Fig. 2). These compounds and six 2-aminothiazole-4-carboxylate

### Ntz analogues



**Fig. 2** Structures of Ntz analogs of Series A and B

derivatives, previously obtained by us [41, 45, 47], were evaluated against *P. falciparum* 3D7. Compounds of A series do not present the nitro substituent in the thiazole ring in order to study the influence of that group in the antimalarial activity.

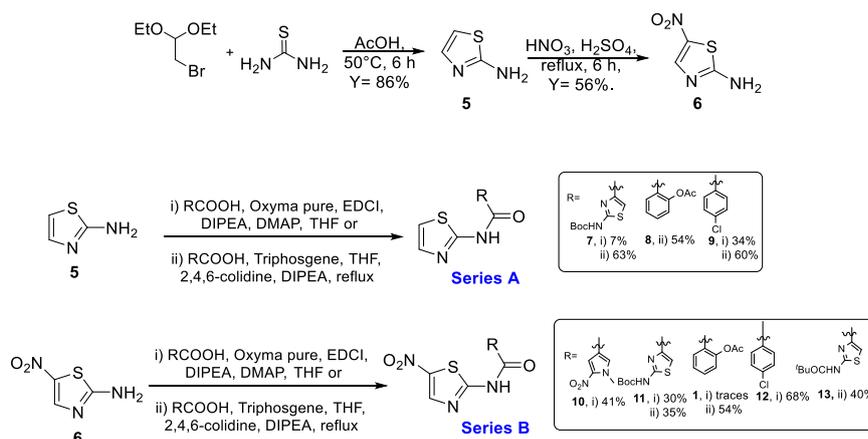
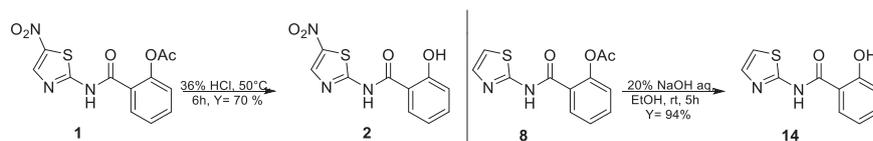
## Results and discussion

### Analog synthesis

The synthesis was started with thiazole preparation from thiourea and bromoacetaldehyde diethyl acetal employing Hanstzch reaction to obtain compound 5, Scheme 1 [48]. Different ratios of EtOH and AcOH were assayed and the best yield was obtained with AcOH 100%. Nitration using Dickey et al. methodology [49], resulted in compound 6 in good yield.

Aware of the low nucleophilicity of the aromatic amine of 2-aminothiazole [41, 47], and consequently, of the low or moderate yield for preparation of the corresponding amide, we paid special attention to the reported methodologies capable of performing this reaction.

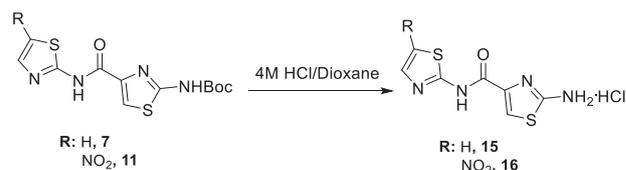
Several groups reported the synthesis of Ntz analogs using acyl chloride derivatives; in other cases EDCI and HOBt or aminocarbonylation followed by iodobenzene substituted reaction were used. All of the methods reported moderate to good yields [50–53]. Recently, our group reported the successful use of oxyma (ethylcianoacetate oxime), in combination with DIC for solid phase peptide

**Scheme 1** Synthesis of Ntz and analogs of type A and B**Scheme 2** Hydrolysis of the ester group of Ntz (**1**) and analog **8**

synthesis (SPPS) or with EDCI for solution macrocyclization [54]. Moreover, Albericio et al. described the impressive coupling efficiency of this coupling agent surpassing HOAT in more demanding peptide models [55]. In addition, the in situ acyl chloride formation using triphosgene is a simple and effective methodology that allowed amide bond formation using aromatic amines [54]. Taking into account these previous results, we decided to explore the use of oxyma/EDCI or triphosgene to obtain amides from 2-aminothiazole compounds **5** or **6**, Scheme 1. The thiazole and pyrrole carboxylic acids used to obtain **7**, **10**, **11**, and **13** were prepared as was described by our group [41].

Compounds **7** and **9** of A series, Scheme 1, were obtained using oxyma/EDCI in low or moderate yield, respectively. As analog **8** was obtained in a higher yield using triphosgene methodology, we explored this methodology for the synthesis of **7** and **9**. Thus, using triphosgene, the yields for these amides derivatives were improved, from 7 to 63% for **7** and from 34 to 60% for **9**. In the case of compounds of B series, Scheme 1, the use of oxyme allowed to obtain **10** and **12** in good (41%) and very good yield (68%), respectively, considering that **12** was obtained by Ballard and co-workers [53], using 4-chlorobenzoyl chloride in 57%. It is important to note that in our case the reaction mixture of **12** was stirred for 4 h at reflux and then at room temperature for 24 h.

Amides **11** and **1** were prepared by both strategies, (i) and (ii) Scheme 1. In the case of **11**, the results were similar, but Ntz (**1**) was obtained only in traces from the reaction mixture using oxyma/EDCI. Nevertheless, the corresponding activated acetylsalicylic acid with oxyma was isolated and its structure was confirmed by NMR and MS analysis.

**Scheme 3** Deprotection of compounds **7** and **11**

In contrast, using triphosgene Ntz was obtained in 54% yield. Taking into account these results, the strategy (ii) was selected to obtain compound **13** in 40% yield based on the recovered starting material.

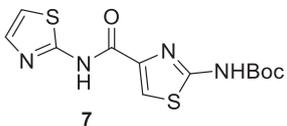
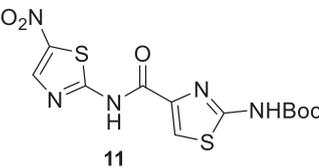
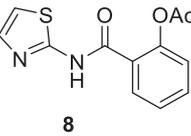
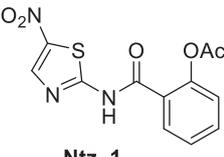
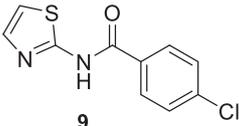
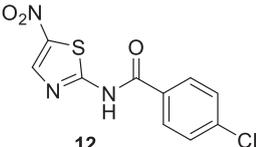
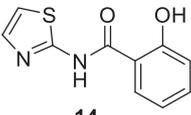
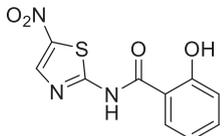
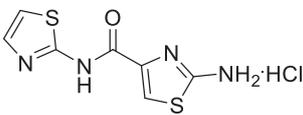
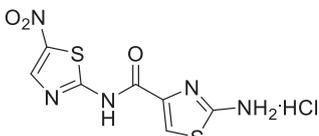
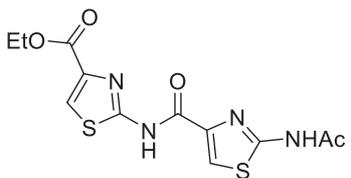
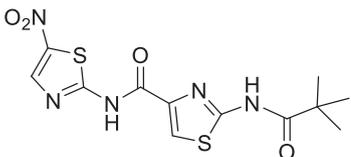
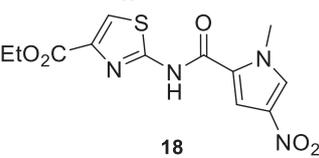
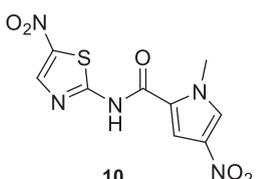
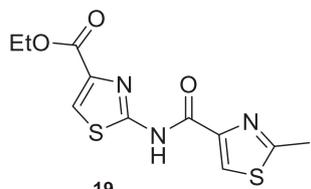
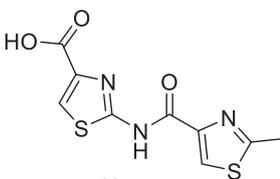
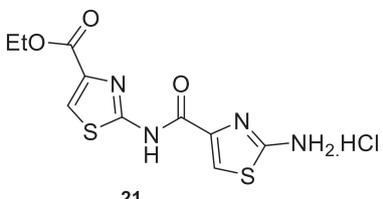
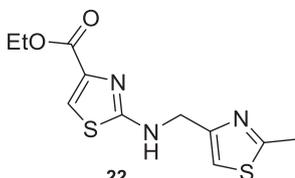
In order to obtain the Ntz metabolite, (**2**, Tiz), first, we tried hydrolysis using basic conditions, 20% NaOH/EtOH (1:1) or 20% LiOH/ EtOH (1:1), but instead of the desired compound, salicylic acid and acetylsalicylic acid were isolated from the reaction mixture. These results allowed us to conclude that in those conditions, the amide bond of **1** is labile. To overcome this difficulty, we intended acid conditions to hydrolyze **1**. Fortunately, the use of HCl 36% at 50 °C during 6 h rendered Tiz (**2**) in 70% yield, Scheme 2. In the case of the Ntz analog (**8**), which does not present a nitro group, the hydrolysis was performed using basic conditions to obtain **14** in excellent yield (94%).

In addition, two derivatives of **7** and **11** were prepared by deprotection of Boc with HCl(g)/Dioxane to obtain **15** and **16**, Scheme 3.

## Antimalarial evaluation

The synthesized compounds and six analogs previously obtained by our group (**17–22**, Table 1) [41, 45, 47] were

**Table 1** Inhibitory activity average (%) at 10  $\mu$ M against *P. falciparum* 3D7, chloroquine-sensitive

Entry	Compounds	Growth reduction average (%)	Entry	Compounds	Growth reduction average (%)
1	 7	5 $\pm$ 2	10	 11	88 $\pm$ 5
2	 8	5 $\pm$ 4	11	 <b>Ntz, 1</b>	30 $\pm$ 10
3	 9	3.2 $\pm$ 0.4	12	 12	34 $\pm$ 9
4	 14	1 $\pm$ 1	13	 <b>Tiz, 2</b>	4 $\pm$ 6
5	 15	7 $\pm$ 9	14	 16	4 $\pm$ 5
6	 17	10 $\pm$ 5	15	 13	85 $\pm$ 1
7	 18	27 $\pm$ 4	16	 10	20 $\pm$ 6
8	 19	6 $\pm$ 7	17	 20	0.6 $\pm$ 0.9
9	 21	23 $\pm$ 1	18	 22	20 $\pm$ 10

assayed at 10  $\mu\text{M}$  for inhibitory activity against the chloroquine-sensitive 3D7 strain of *P. falciparum* using the SYBR Green assay [56]. The inhibitory activities in Table 1 are the average values determined from two independent experiments.

The amides of 2-aminothiazole and the metabolite Tiz (2) showed poor inhibitory activity at 10  $\mu\text{M}$  (% inhibition <5%). The same results were obtained for the derivatives of 2-aminothiazole-4-carboxylate, compounds 17, 19–22. However, some 5-nitro-2-aminothiazole derivatives (10–13) and compound 18 containing 1-methyl-4-nitropyrrole showed moderate growth inhibition of *P. falciparum* 3D7 at 10  $\mu\text{M}$  (% inhibition >27%), suggesting that the presence of the NO<sub>2</sub> substituent was favorable for inhibitory activity. Ntz (1) exhibited moderate activity against *P. falciparum* at 10  $\mu\text{M}$  (% inhibition = 30%), whereas Tiz (2) was a poor inhibitor of *P. falciparum* 3D7 strain (% inhibition = 4%). By contrast, Ntz and Tiz showed IC<sub>50</sub> values in the low micromolar range against *P. berghei* (3.9 and 5.2  $\mu\text{M}$ , respectively) [33]. The parasites' susceptibility to Ntz and Tiz might be related to the genomic and proteomic differences between the *P. falciparum* (human malaria parasite) [57], and *P. berghei* (rodent parasite) [58], species. Indeed, a comparative analysis of the complete proteomes of primate- and rodent-*Plasmodium* species, including *P. falciparum* and *P. berghei*, identified 30 proteins well conserved in the primate parasites that are putatively absent in rodent parasites [59].

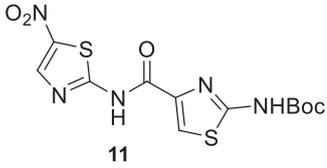
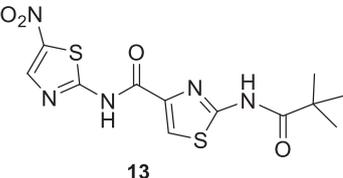
The Boc carbamate 11 and pivalic amide 13, exhibited greater inhibitory activity, 88% and 85% at 10  $\mu\text{M}$ , respectively. In contrast, the amine derivative 16 showed decreased effect against *P. falciparum* 3D7 strain. Compounds 11 and 13 were tested in twofold serial dilutions, in two independent experiments. Table 2 displays the assessed

EC<sub>50</sub> values for 11, 13, and artesunate, tested in parallel as a positive control. As both compounds 11 and 13 showed EC<sub>50</sub> values in the low micromolar range, 5.9 and 8  $\mu\text{M}$ , respectively, they were submitted to cytotoxic assessment against HepG2 cells (hepatocyte carcinoma) and the selectivity indexes (SI) were calculated. In the case of 11, the cytotoxicity determination was limited by the solubility of the compound under the assay conditions (25  $\mu\text{M}$ ). By contrast, compound 13 had the cytotoxic effect assessed (IC<sub>50</sub> = 81  $\mu\text{M}$ ), thereby allowing the determination of the SI. The SI value of 10 indicates a reasonable selectivity for the parasite cells (Table 2).

To further investigate the antiplasmodial activity of the nitazoxanide series, the cross-resistance profile of the representative nitazoxanide analog 11 was assessed against a panel of drug-resistant parasites (Table 3). The panel consisted of representative resistant strains including Dd2, TM90C6B, and 3D7<sup>R</sup>\_848. Dd2 is a chloroquine-, cycloguanil-, and pyrimethamine-resistant strain, while the TM90C6B strain shows resistance against atovaquone. The 3D7<sup>R</sup>\_848 strain was generated in our laboratory and is resistant to MMV692848, a potent *Pf*PI4K inhibitor. To verify the resistant profile of the selected strains, standard antimalarial drugs (artesunate, pyrimethamine, and atovaquone) and MMV692848 were assessed and used as positive controls for inhibition. The pyrimethamine, atovaquone, and MMV692848 showed greater EC<sub>50</sub> values against the Dd2, TM90C6B, and 3D7<sup>R</sup>\_848 strains with resistance indexes (RI) of >222, >1250, and 20, respectively (Table 3). By contrast, the strains in the panel were sensitive to artesunate (EC<sub>50</sub>s = 7–12 nM), confirming their susceptibility to inhibition by small molecules.

The inhibitory activity assessment of 11 against Dd2 and TM90C6B strains indicated that the EC<sub>50</sub> values were three

**Table 2** Half-maximal inhibitory concentration (EC<sub>50</sub>) against *P. falciparum* 3D7, cytotoxicity against HepG2 and selectivity index

Compound	EC <sub>50</sub> <sup>3D7</sup> ( $\mu\text{M}$ ) ± SD	IC <sub>50</sub> <sup>HepG2</sup> ( $\mu\text{M}$ )	Selectivity Index IC <sub>50</sub> <sup>HepG2</sup> / EC <sub>50</sub> <sup>Pf3D7</sup>
 11	5.9 ± 0.7	>25	>4
 13	8 ± 2	81	10
Artesunate	0.013 ± 0.006	-----	----

Values represent the mean ± SD ( $n = 2$ )

**Table 3** Half-maximal inhibitory concentration ( $EC_{50}$ ) against *P. falciparum* sensitive and resistance strains, resistance index (RI =  $EC_{50}^{\text{resistant}} / EC_{50}^{3D7}$ )

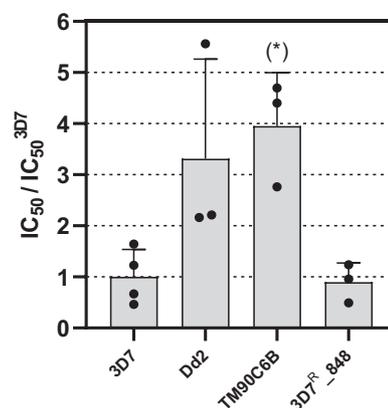
Compound	$EC_{50}$ ( $\mu\text{M}$ ) average $\pm$ SD							
	3D7	Dd2	RI	TM90C6B	RI	3D7 <sup>R</sup> _848	RI	
11	8 $\pm$ 4	30 $\pm$ 20	3.8	31 $\pm$ 8	3.9	7 $\pm$ 3	0.9	
Artesunate	0.008 $\pm$ 0.002	0.007 $\pm$ 0.003	0.9	0.0069 $\pm$ 0.0006	0.9	0.012 $\pm$ 0.006	1.5	
Pyrimethamine	0.045 $\pm$ 0.004	>10	>222	–	–	–	–	
Atovaquone	0.0008 $\pm$ 0.0002	–	–	>1	>1250	–	–	
MMV692848	0.12 $\pm$ 0.02	–	–	–	–	2.4 $\pm$ 0.3	20	

and fourfold greater, respectively than the  $EC_{50}$  values against 3D7 sensitive strain, while the  $EC_{50}$  value against 3D7<sup>R</sup>\_848 strain was similar to the  $EC_{50}$  value against the sensitive strain (Fig. 3). These findings indicate that **11** has comparable potency against the resistant strains in the panel. Moreover, the RI values varied from 0.9 to 3.9 (Table 3), which were considerably smaller than the RI values observed for the standard antimalarials against the resistant strains (RI = 20 – >1250). These findings suggest no cross-resistance between the bis thiazole derivative (**11**) and the antimalarials pyrimethamine, atovaquone, and MMV692848.

The main molecular target of nitazoxanide in protozoa and anaerobic bacteria is pyruvate:ferredoxin oxidoreductase (PFOR), a biomolecule involved in the enzyme-dependent electron transfer reaction and essential to the anaerobic energy metabolism, especially, in anaerobic protozoan parasites [60, 61]. Nevertheless, to the best of our knowledge, no genes encoding PFOR have been isolated from *P. falciparum*, suggesting that the antiplasmodial activity of the nitazoxanide derivatives is via other mechanism of action. In line with this, organisms that lack the PFOR and resistant to metronidazole (a nitroimidazole derivative that targets PFOR catalytic reaction) are sensitive to nitazoxanide and nitazoxanide derivatives [62, 63]. Furthermore, compound **11** showed comparable potencies against the sensitive and resistant strains of the parasite, indicating that the mechanism of action underlying the antiplasmodial activity is not related to the inhibition of the dihydrofolate reductase (DHFR), cytochrome bc1 complex (cyt *bc1*), and phosphatidylinositol 4-OH kinase (PI4K) enzymes. Therefore, the mechanism of action of **11** against *P. falciparum* is unknown and shall be investigated.

## Conclusion

Two methodologies were studied to prepare NTz and analogs. The use of triphosgene rendered higher yield than the use of oxyma to obtain the desired amides. All the compounds of A series and all the amide derivatives of 2-aminothiazole-4-carboxylate, which not present nitro group showed poor inhibitory activity against *P. falciparum*



**Fig. 3** Half-maximal inhibitory concentration of **11** against sensitive (3D7) and resistant (Dd2, TM90C6B and 3D7<sup>R</sup>\_848) strains, normalized by the  $IC_{50}$  value against the sensitive strain (\* $P < 0.05$ )

(3D7 strain) at 10  $\mu\text{M}$ . Ntz showed moderate inhibition with 30% of growth reduction of parasite at 10  $\mu\text{M}$  and Tiz was a poor inhibitor of *P. falciparum* 3D7 strain. As the previous evaluation of Ntz and Tiz, carried out by Navarrete-Vazquez et al. [33], was performed against schizonts of the rodent *P. berghei*, our investigations unveiled weak activity for Ntz against the human parasite *P. falciparum*. However, the bis thiazole analogs (**11** and **13**), showed attractive inhibitory activity with  $EC_{50}$  values of 5.9  $\mu\text{M}$  and 8  $\mu\text{M}$ , and low propensity to show toxic effect against HepG2 cells. Moreover, the representative compound of the series (**11**) showed no cross-resistance against a panel of sensitive and resistant parasites. We concluded that the nitro group of **11** and **13** play an important role for their bioactivity against the human malaria parasite. Taking into account that the amine derivative **16** is a poor inhibitor, it could be concluded that the presence of an amide or carbamate in the second thiazole ring, derived from 2-aminothiazole-4-carboxylate, improve the antimalarial activity.

## Experimental

Most of the reagents were purchased from commercial suppliers and used without further purification, some of them were synthesized by part of the group.

$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded on Bruker Avance Neo 400 MHz spectrometer using the indicated deuterated solvents and TMS as an internal standard. Chemical shifts ( $\delta$ ) are given in parts per million (ppm), multiplicities according to: singlet (s), broad singlet (ba), doublet (d), triplet (t), (doublet of doublets (dd), doublet of triplets (dt), doublet of doublet of doublets (ddd), multiplet (m). Coupling constants ( $J$ ) are expressed in Hz.

All mass spectra were acquired with a Shimadzu 8040 HPLC-MS-MS equipment, with LC-20AD pumps, a SIL-20A autosampler, electrospray ionization and triple-quadrupole mass detector. Flash column chromatography was carried out with silica gel 60 (J.T. Baker, 40  $\mu\text{m}$  average particle diameter). All reactions and chromatographic separations were monitored by TLC, conducted on 0.25-mm silica gel plastic sheets (Macherey/Nagel, Polygram\_SIL G/UV 254). TLC plates were analyzed under 254 nm UV light, iodine vapor, or ninhydrine spray. Yields are reported for chromatographically and spectroscopically ( $^1\text{H}$  and  $^{13}\text{C}$  NMR) pure compounds.

All solvents were purified according to literature procedures [64]. All reactions were carried in dry, freshly distilled solvents under anhydrous conditions unless otherwise stated.

### General procedure for amide bond formation using oxyma pure

Oxyma pure (1.2 mmol), EDCI (1.2 mmol), DIPEA (1.2 mmol), 4-DMAP (catalytic) were added to a stirred solution of the respective acid (1.2 mmol) and amine (1.0 mmol) under  $\text{N}_2$  atmosphere in dry solvent ( $\text{CH}_2\text{Cl}_2$  or THF) at  $0^\circ\text{C}$ . The resulting mixture was stirred at room temperature for 12–48 h and then evaporated in vacuo. The crude was dissolved in EtOAc, washed with 5% v/v HCl and with a saturated solution of  $\text{NaHCO}_3$ , dried with  $\text{Na}_2\text{SO}_4$ , filtered, and evaporated in vacuo. The crude was purified by flash chromatography using the corresponding eluent to give the amide.

### General procedure for amide bond formation using triphosgene

To a stirred solution of (1.0 mmol) acid, (0.4 mmol) triphosgene were added in dry THF under  $\text{N}_2$  atmosphere at room temperature. After 10 min 2.4 mmol of 2,4,6-colidine were added, and a white precipitate was formed. After 30 min, 0.9 mmol of the respective amine and 0.9 mmol of DIPEA were added. The resulting mixture was stirred at reflux for 12–24 h. The mixture is concentrated to dryness and the crude was dissolved in EtOAc, washed with 5% v/v HCl and a saturated solution of  $\text{NaHCO}_3$ , dried with  $\text{Na}_2\text{SO}_4$ , filtered and evaporated in vacuo. The crude was

purified by flash chromatography using the corresponding eluent to give the amide.

### 2-[(5-Nitro-1,3-thiazol-2-yl)carbamoyl]phenyl acetate (1)

Compound **1** was obtained following the general procedure for in situ acid chloride formation from **6** (236 mg, 1.63 mmol) and acetylsalicylic acid, in 19 mL of dry THF. Reaction was refluxed 6 h and stirred 14 h at  $40\text{--}50^\circ\text{C}$ . Yield 42%. Brown powder.  $R_f = 0.35$  using a mixture of  $\text{CHCl}_3\text{:MeOH}$  (3:0.2) as eluent.  $^1\text{H}$  NMR (400 MHz, DMSO)  $\delta$  (ppm): 13.61 (ba, 1H), 8.68 (s, 1H), 7.87 (dd,  $J = 7.9, 1.6$  Hz, 1H), 7.67 (td,  $J = 7.9, 1.7$  Hz, 1H), 7.43 (td,  $J = 7.6, 1.0$  Hz, 1H), 7.30 (dd,  $J = 8.17, 0.9$  Hz, 1H), 2.26 (s, 3H).  $^{13}\text{C}$  NMR (100 MHz, Acetona- $d_6$ )  $\delta$  (ppm): 168.7, 164.7, 162.3, 151.3, 148.9, 141.4, 133.7, 129.7, 126.1, 124.2, 123.7, 20.0. MS (ESI+) calcd for  $\text{C}_{12}\text{H}_9\text{N}_3\text{O}_5\text{S}$ ,  $[\text{M} + \text{H}]^+$  308.02, found 308.05.

### tert-Butyl 4-((5-nitrothiazol-2-yl)carbamoyl)thiazol-2-yl) carbamate (11)

Compound **11** was obtained following the general procedure for amide bond formation using oxyma pure from **6** (80 mg, 0.55 mmol) and 2-((tert-butoxycarbonyl)amino)thiazole-4-carboxylic acid in 8 mL of dry THF. Reaction was refluxed 3 h and stirred 14 h at  $40\text{--}50^\circ\text{C}$ . Yield 35%. White powder.  $R_f = 0.24$  with a mixture of  $\text{CHCl}_3\text{:MeOH}$  (3:0.1) as eluent.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 10.51 (s, 1H), 8.38 (s, 1H), 8.0 (s, 1H), 3.91 (s, 1H), 1.58 (s, 9H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 176.4, 162.3, 160.8, 160.6, 146.4, 141.3, 141.0, 121.8, 84.2, 28.1. MS (ESI-)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{13}\text{N}_5\text{O}_5\text{S}_2$   $[\text{M} - \text{H}]^-$  370.03, found 370.10.

### SYBR green I growth inhibition assay against *P. falciparum* asexual forms

Continuous in vitro cultures of *P. falciparum* (strains 3D7, TM90C6B, and 3D7<sup>R</sup>\_848) were kept using an adaptation of the method described by Trager and Jansen [65]. *P. falciparum* cultures were synchronized for assays by treatment with a solution of 5% sorbitol for 10 min, as described by Lambros and Vanderberg [66]. Compounds were diluted to a stock concentration of 20 mM in 100% DMSO before the experiments and maintained at  $-20^\circ\text{C}$ . Compound inhibitory potencies were determined using the SYBR Green I phenotypic assay (Smilkstein et al.) [67]. Briefly, in 96-well plates, 180  $\mu\text{L}$  of a parasite suspension in the early trophozoite (ring) form at 0.5% parasitemia and 2% hematocrit was incubated for 72 h with 20  $\mu\text{L}$  of 10 $\times$  concentrated serial dilutions of each compound. The anti-malarial artesunate, pyrimethamine, atovaquone, and

MMV692848 were used as positive controls of inhibition for the cross-resistance experiments. The plates were kept at 37 °C and in a low oxygen atmosphere (5% O<sub>2</sub>, 5% CO<sub>2</sub>, 90% N<sub>2</sub>). Positive and negative growth controls were added to each independent plate. The growth medium was then removed, and the deposited RBCs were resuspended in PBS buffer (116 mM NaCl, 10 mM Na<sub>2</sub>HPO<sub>4</sub>, 3 mM KH<sub>2</sub>PO<sub>4</sub>). A solution of SYBR Green I DNA Stain diluted was added in a lysis buffer (20 mM Tris, 5 mM EDTA, 0.008% (m/v) saponin, and 0.08% (m/v) Triton X-100, at pH 7.5) to induce hemolysis. The plates were incubated for an additional 30 min, after which the fluorescence of the plate was measured (absorption and emission wavelengths of 485 nm and 535 nm, respectively). Fluorescence intensity was analyzed in terms of parasite viability as compared to controls, using the Origin 9.0 software (OriginLab, Northampton, MA, USA). Concentration-response curves were built, and half-maximal inhibitory concentration (IC<sub>50</sub>) values were determined for each compound using nonlinear regression analysis.

#### Cultivation of human hepatocellular carcinoma cells (HepG2 cell line)

A culture of HepG2 cells was kept in a flask in a humidified atmosphere of 5% CO<sub>2</sub> at 37 °C. The culture medium used was RPMI 1640 supplemented with 25 mM HEPES (pH 7.4), 24 mM sodium bicarbonate, 11 mM D-glucose, 40 µg/mL penicillin-streptomycin, and 10% (v/v) bovine fetal serum. Every 3–4 days, treatment with a 0.25% trypsin solution was used to release cells from the flask walls and a 1:4 proportion of the cells were maintained in culture.

#### MTT assay for cytotoxicity evaluation

An adaptation of the MTT assay described by Denizot and Lang [68] was employed to determine cytotoxic activity. The HepG2 cells were counted and distributed in a 96-well plate, in a proportion of 5 × 10<sup>6</sup> cells per well. The plate was incubated overnight at 37 °C, in a humidified atmosphere of 5% CO<sub>2</sub>. After that, serial dilutions of the compounds were added in a 1:9 proportion to each well, and the plates were incubated for another 24 h. Positive (no compound) controls were added to each plate for normalization of results. The supernatant was then removed, and a solution of MTT salt (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) was added to each well. After 2–4 h of incubation, the formazan crystals formed by MTT reduction were solubilized in DMSO. The absorbance of the plate at 570 nm was measured, and the intensity values obtained were converted to viability values using the equation below. Dose-response curves were crafted for each compound using the OriginPro 9.0 software (OriginLab), and the minimum inhibitory

concentration for 50% of cells (IC<sub>50</sub><sup>HepG2</sup>) was determined for each compound. The selectivity index (SI) was calculated by the ratio of IC<sub>50</sub><sup>HepG2</sup> to IC<sub>50</sub><sup>3D7</sup>.

#### Data availability

NMR and MS spectra are depicted in the Supplementary Material.

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#### Compliance with ethical standards

**Conflict of interest** The authors declare no competing interests.

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